

A new species of *Rhyacophila* Pictet, 1834 (Trichoptera, Rhyacophilidae) from Corsica with the genomic characterization of the holotype

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Abstract

We describe a new species in the *Rhyacophila tristis* group, *Rhyacophila lignumvallis* Graf & Rázuri-Gonzales, **sp. nov.**, from the island of Corsica (France) based on a single male specimen. In addition to the morphological differences between the new species and the most similar species in the group, we also provide a phylogenetic tree based on the mitochondrial cytochrome c oxidase subunit I (mtCOI), including sequences from 16 out of the 28 currently recognized species in the group. These data, together with conspecific larval sequences, support the status of the new species and shed light on an additional potential new species near *Rhyacophila pubescens*. Using a low-cost next-generation sequencing approach, we generated the mito- and draft nuclear genome assembly of the holotype of *R. lignumvallis* **sp. nov.** as well as that of *R. tsurakiana*. This genetic data represents an important additional characterization to the description of morphological features and is valuable for future investigations, such as population or phylogenomic studies.

Key words: Caddisflies, holotype genomics, mitogenome, taxonomy



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Introduction

The genus *Rhyacophila* Pictet 1834, with 814 extant and 30 fossil species, is the largest caddisfly genus in the world (Valladolid et al. 2023). These caddisflies are primarily distributed in the northern hemisphere, but they also occur in temperate and tropical India and southeastern Asia (Holzenthal et al. 2007). Given the sheer size of the genus, many species groups have been proposed based on the morphological characteristics of the larvae (Döhler 1950) and the male genitalia (Ross 1956; Schmid 1970). However, the groups and subgenera proposed by different authors do not correspond to each other. For example, some of the larval characters for the subgenera of Döhler are found in several of the groups of Schmid. Phylogenetic studies of different groups proposed by

Schmid (1970) seem to showcase the overall utility of his system, even if rearrangements are sometimes needed (Coppa et al. 2012).

One of the groups proposed by Schmid is the *Rhyacophila tristis* group in the *R. invaria* branch. It is mainly characterized by a large segment IX without an apical dorsal lobe, a simple and oblique segment X, with fairly large anal sclerites, which can be joined or partially fused to each other and fused to segment X, very large phallotheca with a simple or complex dorsal arm, a simple chitinous aedeagus, simple lobe-like parameres, and lacking a ventral lobe, among other characteristics (Schmid 1970). Currently, the species group contains 28 species in two subgroups: the *tristis* and the *pubescens* subgroups (Schmid 1970, Suppl. material 1). The *tristis* subgroup is further characterized by the complex shape of the dorsal arm of the phallotheca and the presence of fairly large parameres fused to the aedeagus. The *pubescens* subgroup, on the other hand, has a simpler dorsal arm of the phallotheca and elongated free parameres. Species of the *R. tristis* species group (i.e., both subgroups) generally occur in headwaters to medium-sized, fast-flowing streams at middle elevations throughout Europe (excluding the British Isles, Northern Europe, and Russia) and Western Asia (i.e., Turkey and Iran) (Schmid 1970; Coppa et al. 2012; Suppl. material 1).

Only two species in the *R. tristis* group were previously known on the island of Corsica (France): *R. pubescens* Pictet, 1834 and *R. tristis* Pictet, 1834. Both species were initially recorded by Mosely (1930, 1932). More recently, Engelhardt (2009) assessed the phylogenetic relationships in the *R. tristis* group and the phylogeography of *R. pubescens* throughout its distributional range. Engelhardt showed that the larval specimens of *R. pubescens* from the island were significantly divergent from all the other populations. Moreover, the Corsican specimens formed a highly supported clade, separate from all other *R. pubescens* (Engelhardt et al. 2011).

In the present paper, we confirm the genetic and morphological distinctness of adult males of the Corsican lineage to represent a new species and describe it as *Rhyacophila lignumvallis* sp. nov. in the *pubescens* subgroup of the *R. tristis* group. Additionally, we present an annotated draft genome from the holotype and its complete mitogenome, adding valuable genetic information to the holotype description. Finally, we also include a draft genome and the mitogenome for a second species in the *Rhyacophila tristis* species group, *R. tsurakiana* Malicky, 1984 from Albania. We hope these genomic resources stimulate research on this group of insects, especially since their diversity is high in this area and they seem to be restricted to specific mountain ranges, as suggested by Oláh et al. (2022).

Material and methods

Specimen collection

The specimen was swept from the riparian vegetation using a hand net.

DNA extraction, library preparation, and whole genome sequencing

A pair of legs from the holotype of *R. lignumvallis* sp. nov. were removed, and the tissue was incubated overnight in 60 µl TNES lysis buffer (100 mM Tris-HCl, 25 mM NaCl, 10 mM EDTA, 1% SDS) and 8 µl Proteinase K (20 mg/ml). For DNA

binding and cleanup, 75 µl custom speed-bead suspension was added (Sera-Mag SpeedBeads Carboxylate, hydrophobic, Cytiva; see Rohland and Reich 2012), incubated for 15 min on a rotating shaker, and the beads were washed twice with 75% ethanol after the supernatant had been removed and discarded. The DNA was eluted in 1X TE.

DNA sequences were generated for the cytochrome c oxidase subunit I barcoding region (mtCOI, 658 bp) using primers LCO1490-L and HCO2198-L (Nelson et al. 2007). Polymerase chain reactions (PCR) were run on a Mastercycler Pro S (Eppendorf, Hamburg, Germany) in reactions containing 1X MyTaq Reaction Buffer, 0.4 µM of each forward and reverse primer, 0.5 U MyTaq DNA Polymerase, 1 µl DNA and nuclease-free water to fill up to a total volume of 10 µl. Reaction conditions were 1 min at 95 °C for initial denaturation followed by 35 cycles of 20 s at 95 °C (denaturation), 30 s at 45 °C (annealing), and 30 s at 72 °C (extension). The reaction ended with a final extension for 5 min at 72 °C. PCR products were visualized on agarose gels and purified using a modified ExoSAP protocol with Exonuclease I (20U/µl) and Fast AP Thermoinsensitive Alkaline Phosphatase (1U/µl; both ThermoFisher Scientific, Vilnius, Lithuania). DNA sequences were generated at the Laboratory Centre of the Senckenberg Biodiversity and Climate Research Centre using a 3730XL DNA Analyzer (Applied Biosystems).

Genomic DNA was taken from the above DNA isolates, quantified using a Qubit 4.0 fluorometer with a 1x dsDNA HS Assay Kit (ThermoFisher Scientific, Waltham, USA), and sheared to a target fragment size of 350 bp using a Bioruptor Pico (Diagenode, Seraing, Belgium). Genomic libraries were prepared from 27.4 ng sheared gDNA using the NEBNext Ultra II DNA Library Preparation Kit for Illumina (New England Biolabs, Ipswich, MA, USA), following the manufacturer's manual. Adapters were diluted 1:10 as recommended for low input libraries, and size selection was omitted due to the low DNA amount. A dual indexing PCR was run for 7 cycles on a Mastercycler (Eppendorf, Germany) using NEBNext Multiplex Oligos for Illumina (Dual Index Primers Set 1; New England Biolabs, Ipswich, MA, USA). After cleanup, library integrity was verified on a 2200 TapeStation with a High Sensitivity D1000 Tape (Agilent, Santa Clara, CA, USA), and shipped for 150 bp paired-end sequencing (ordering 30 Gbp output) on a partial lane of an Illumina NovaSeq 6000 platform (San Diego, CA) at Novogene (Cambridge, UK).

Raw reads are deposited in the National Center for Biotechnology Information's Sequence Read Archive (NCBI SRA) under the accession number SRR22799047 under Bioproject PRJNA899095.

DNA barcoding and phylogenetic analysis

The final mtCOI alignment included 71 sequences: 68 sequences from 16 species and 2 morphospecies in the *Rhyacophila tristis* group. *Rhyacophila italica* Moretti, 1981, *Himalopsyche kuldschensis* (Ulmer, 1927), and *H. triloba* (Hwang, 1958) were included as outgroups. All sequences were generated for this manuscript, except *R. bosnica* Schmid, 1970 ([MK211322](#)), *H. kuldschensis* ([KX143534](#)), and *H. triloba* ([KX295339](#)), which were retrieved from GenBank. The barcode region is 658 bp in length. However, some of our sequences were incomplete, and their lengths were between 576 and 658 bp. Therefore, the final alignment was completed with Ns on both ends, reaching 3.69% of missing data. Sequence specimen data and GenBank accession numbers are summarized in Table 1.

Table 1. Sequence specimen data, with GenBank accession numbers, of the studied *Rhyacophila* species and the out-groups *Himalopsyche kuldschensis* (Ulmer, 1927) and *H. triloba* (Hwang, 1958).

Species	Country*	Locality	Latitude, Longitude	Accession No.
<i>Himalopsyche kuldschensis</i>	KG	Kalay Makhmud valley between Or-Mazan-Suu and Ala Malden	39.683, 70.8833	KX143534
<i>Himalopsyche triloba</i>	CN	Sichuan, near Jiuzhaigou	30.45, 102.50	KX295339
<i>R. akutila</i>	BG	Prava Marica stream at Zavratica mountain hut	42.16789, 23.64139	PP515197
<i>R. aquitanica</i>	FR	Ruisseau de Chousse, upper tributary; between Arrette & La Pierre Saint-Martin	43.00757, -0.73572	PP515198
<i>R. aquitanica</i>	FR	Ruisseau de Chousse, upper tributary; between Arrette & La Pierre Saint-Martin	43.00757, -0.73572	PP515199
<i>R. aquitanica</i>	FR	Ruisseau de Chousse, upper tributary; between Arrette & La Pierre Saint-Martin	43.00757, -0.73572	PP515200
<i>R. aquitanica</i>	FR	Ruisseau de Chousse, upper tributary; between Arrette & La Pierre Saint-Martin	43.00757, -0.73572	PP515201
<i>R. aquitanica</i>	ES	tributaries to the Barranco de Urdiceto, above Embalse de Urdiceto	42.67832, -0.2772	PP515202
<i>R. bosnica</i>	BA	Vareš municipality, Rajčevački stream	–	MK211322
<i>R. carpathica</i>	RO	Galeş Lake	45.38650, 22.90914	PP515204
<i>R. carpathica</i>	RO	Galeş Lake	45.38650, 22.90914	PP515205
<i>R. carpathica</i>	RO	Galeş Lake	45.38650, 22.90914	PP515206
<i>R. carpathica</i>	RO	Galeş Lake	45.38650, 22.90914	PP515207
<i>R. carpathica</i>	RO	Caraş-Severin, Iauna Mare stream	45.51636, 22.59017	PP515208
<i>R. carpathica</i>	RO	Caraş-Severin, Poiana Mărului	45.39583, 22.53422	PP515209
<i>R. cibinensis</i>	RO	unnamed stream near Păltiniş	45.63878, 23.92540	PP515210
<i>R. cibinensis</i>	RO	unnamed stream near Păltiniş	45.63878, 23.92540	PP515211
<i>R. cibinensis</i>	RO	unnamed stream near Păltiniş	45.63878, 23.92540	PP515212
<i>R. cibinensis</i>	RO	Lotru river	45.38, 23.62	PP515213
<i>R. italica</i>	IT	Purello	43.32, 12.77	PP515214
<i>R. lignumvallis</i> sp. nov.	FR	Corsica, Tributary to the Tavignano	42.25639, 9.20583	PP515216
<i>R. lignumvallis</i> sp. nov.	FR	Corsica, Tributary to the Tavignano	42.25639, 9.20583	PP515217
<i>R. lignumvallis</i> sp. nov.	FR	Corsica, Tributary to the Tavignano	42.25639, 9.20583	PP515218
<i>R. lignumvallis</i> sp. nov.	FR	Corsica, bridge over the river Vecchio near the confluence with the river Tavignano	42.2275, 9.24306	PP515215
<i>R. margaritae</i>	BG	Lower left tributary to Zavodna, above Ribaritsa village and below Vezhen peak	42.76, 24.37	PP515219
<i>R. margaritae</i>	BG	Lower left tributary to Zavodna, above Ribaritsa village and below Vezhen peak	42.76, 24.37	PP515220
<i>R. margaritae</i>	BG	Lower left tributary to Zavodna, above Ribaritsa village and below Vezhen peak	42.76, 24.37	PP515221
<i>R. obtusa</i>	BG	Zavodna river, upstream of the confluence with the Beli Vit at the Ribaritsa village	42.812, 24.371	PP515222
<i>R. obtusa</i>	BG	Zavodna river, upstream of the confluence with the Beli Vit at the Ribaritsa village	42.791, 24.377	PP515223
<i>R. orghidani</i>	RO	right-side inflow of Leşu artificial lake	46.80981, 22.58948	PP515224
<i>R. orghidani</i>	RO	right-side inflow of Leşu artificial lake	46.80981, 22.58948	PP515225
<i>R. orghidani</i>	RO	Băișoara	46.53287, 23.28078	PP515226
<i>R. orghidani</i>	RO	Băișoara	46.53287, 23.28078	PP515227
<i>R. pirinica</i>	BG	24.5 km NNW from Gotse Delchev	41.63156, 23.44628	PP515228
<i>R. pirinica</i>	BG	24.5 km NNW from Gotse Delchev	41.63156, 23.44628	PP515229
<i>R. pirinica</i>	BG	24.5 km NNW from Gotse Delchev	41.63156, 23.44628	PP515230
<i>R. producta</i>	AT	Nockberge	46.85, 13.76	PP515231
<i>R. producta</i>	AT	Nockberge	46.85, 13.76	PP515232
<i>R. pubescens</i>	CH	La Motte above Ocourt	47.35, 7.06	PP515233

Species	Country*	Locality	Latitude, Longitude	Accession No.
<i>R. pubescens</i>	CH	La Motte above Ocourt	47.35, 7.06	PP515234
<i>R. pubescens</i>	FR	Ravin de Chambières	43.93278, 6.63694	PP515235
<i>R. pubescens</i>	FR	Ravin de Chambières	43.93278, 6.63694	PP515236
<i>R. pubescens</i>	FR	La Condamine-Châtelard	44.451, 6.741	PP515237
<i>R. pubescens</i>	FR	La Condamine-Châtelard	44.451, 6.741	PP515238
<i>R. pubescens</i>	FR	La Condamine-Châtelard	44.451, 6.741	PP515239
<i>R. pubescens</i>	IT	Tributary of Fiume Tescio	43.09722, 12.67556	PP515240
<i>R. pubescens</i>	IT	Tributary of Fiume Tescio	43.09722, 12.67556	PP515241
<i>R. pubescens</i>	IT	Tributary of Fiume Tescio	43.09722, 12.67556	PP515242
<i>R. pubescens</i>	IT	Nameless brook near Rezzo	44.02583, 7.86667	PP515243
<i>R. pubescens</i>	IT	Valle di Pietra	44.07722, 7.80639	PP515244
<i>R. pubescens</i>	IT	Valle di Pietra	44.07722, 7.80639	PP515245
<i>R. sarplana</i>	AL	Tropojë, open stream on Mt. Callumit, above town	42.49862, 20.12443	PP515203
<i>Rhyacophila</i> sp., <i>tristis</i> grp.	AT	Carinthia, Gail river at Kötschach-Mauthen town	46.67, 12.98	PP515255
<i>Rhyacophila</i> sp., <i>tristis</i> grp.	IT	Lombardia, Valle del Ferro	45.77277, 9.98996	PP515256
<i>Rhyacophila</i> sp., <i>tristis</i> grp.	IT	Trentino-Alto Adige/Südtirol, Camposilvano	45.75988, 11.14189	PP515257
<i>Rhyacophila</i> sp., <i>tristis</i> grp.	FR	Ruisseau de Chousse, upper tributary; between Arrette & La Pierre Saint-Martin	43.00757, -0.73572	PP515258
<i>R. trescavicensis</i>	ME	Ali-pašini springs	42.54706, 19.83240	PP515246
<i>R. trescavicensis</i>	ME	Ali-pašini springs	42.54706, 19.83240	PP515247
<i>R. trescavicensis</i>	ME	Ali-pašini springs	42.54706, 19.83240	PP515248
<i>R. trescavicensis</i>	ME	Ali-pašini springs	42.54706, 19.83240	PP515249
<i>R. tristis</i>	RO	Hunedoara, Câmpu lui Neag	45.30227, 22.97388	PP515250
<i>R. tristis</i>	RO	Hunedoara, Câmpu lui Neag	45.30227, 22.97388	PP515251
<i>R. tristis</i>	RO	Covasna, Comandău	45.81488, 26.32934	PP515252
<i>R. tristis</i>	RO	Harghita, Voșlăbeni	46.6815, 25.6738	PP515253
<i>R. tristis</i>	RO	Vâlcea, Voineasa, Lotru river	45.463, 23.62	PP515254
<i>R. tsurakiana</i>	AL	river Shushica at the village of Brataj	40.26622, 19.67198	PP515259
<i>R. vranitzensis</i>	BA	Sljeme	43.9403, 18.5122	PP515260
<i>R. vranitzensis</i>	BA	Sljeme	43.9403, 18.5122	PP515261
<i>R. vranitzensis</i>	BA	Sljeme	43.9403, 18.5122	PP515262
<i>R. vranitzensis</i>	BA	Skakavac waterfall	43.94238, 18.44196	PP515263
<i>R. vranitzensis</i>	BA	Skakavac waterfall	43.94238, 18.44196	PP515265

* Country codes in the ISO 3166-1 alpha-2 standard

The maximum likelihood tree was produced using IQ-TREE v.2.1.3 (Minh et al. 2020), using the command `iqtree2 -s RhyacophilaLignumvalleMS_658bp.fasta -B 1000 -bnni -alrt 1000 --prefix RhyacophilaLignumvalleMS_658bp`. The TIM2+F+I+G4 nucleotide substitution model was selected using ModelFinder (Kalyaanamoorthy et al. 2017). Statistical support for the tree topology was assessed with the ultrafast bootstrap approximation (UFboot) (Hoang et al. 2017). Clades with UFboot $\geq 95\%$ are considered well-supported. Additionally, each bootstrap tree was optimized with a hill-climbing nearest neighbor interchange (NNI) search (flag -bnni in the command above) based on the corresponding bootstrap alignment to prevent overestimating UFboot branch support values, as recommended by Hoang et al. (2017).

The consensus tree was visualized and edited in TreeViewer v.2.2.0 (Bianchini and Sánchez-Baracaldo 2024). Additional aesthetic edits were made in Adobe Illustrator CS6.

Holotype mitogenome and nuclear genome assembly

After quality control with FastQC v.0.11.9 (Andrews 2019), raw reads were trimmed for low-quality regions, adapter sequences, and overrepresented *k-mers* using autotrim.pl v.0.6.1 (Waldvogel et al. 2018) and Trimmomatic v.0.39 (Bolger et al. 2014) with the adapter_all.fa of Trimmomatic and the following settings ILLUMINACLIP:2:30:10:8:true, SLIDINGWINDOW:4:20, MINLEN:50, and TOPHRED33. Unpaired reads were discarded and paired reads were checked for contamination using Kraken v.2.0.9 (Wood and Salzberg 2014).

Genome size was estimated using a method based on *k-mer* distribution. For this, *k-mers* were counted with JELLYFISH v.2.3.0 (Marçais and Kingsford 2011) using *jellyfish count -C -s 1000000000 -F 2* and a *k-mer* length of 21 based on the raw sequence reads. A histogram of *k-mer* frequencies was created with *jellyfish histo* and used for analysis with the online web tool GenomeScope v.2.0 (Ranallo-Benavidez et al. 2020) using the following parameters: *k-mer* length = 21, ploidy = 2, max *k-mer* coverage = 10000.

The mitochondrial genomes were first assembled with the raw reads using NOVOplasty v.4.2 (Dierckxsens et al. 2016) using the following parameters: type = mito, genome range = 12000–22000, *k-mer* = 33, max memory = 100, read length = 150, insert size = 300, platform = illumina, paired = PE, insert size auto = yes. The partial sequence of the cytochrome c oxidase subunit I (COX1) gene of *Rhyacophila fasciata* Hagen, 1859 ([MT559357.1](#)) was used as seed input. All other parameters were kept as default. In addition, we used a second mitogenome assembler MitoZ v.2.3 (Meng et al. 2019). For this purpose, the raw data was subsampled to 10,000,000 reads using seqk and then used as input for *MitoZ assemble* with the following parameters: genetic_code 5, clade Arthropoda, fastq_read_length 150, insert_size 300, run_mode 2, filter_taxa_method 1, requiring_taxa 'Arthropoda'. Annotation of tRNAs, rRNAs, and protein-coding genes was done for the best mitogenome assembly of each species with MitoZ v.2.3 using the module annotate with genetic_code 5 and clade Arthropoda. Both mitogenome assemblies were aligned to the complete mitogenome of *R. quadrifida* Sun & Yang, 1995 ([OL678049.1](#)) and *R. kando* Schmid, 1970 ([OL678048.1](#)) with MAFFT in Geneious Prime v.2022.1.1 (Biomatters Ltd.) to set the correct start position and manually curate the control-region. The mitochondrial genome assembly was deposited in GenBank under the accession [OQ984043](#).

Nuclear genome assembly was conducted in Spades v.3.14.1 (Bankevich et al. 2012) with the default settings. Scaffolds smaller than 500 bp and those with blast hits to the mitochondrial genome assembly were filtered out. Assembly statistics were calculated with Quast v.5.0.2 (Gurevich et al. 2013) and completeness was assessed via screening for single-copy orthologs with BUSCO v.5.2.2 (Manni et al. 2021) using the endopterygota_odb10 dataset. As an additional quality control, trimmed reads were mapped back to the assembly with bwa-mem v.0.7.17-r1188 (Li 2013) with parameters -a -c 10000, and the back-mapping rate was calculated with qualimap v.2.2.1 (Okonechnikov et al. 2015). To check for potential contamination, taxon-annotated GC-coverage (TAGC) plots were generated with BlobTools v.1.1.1 (Laetsch and Blaxter 2017). For this purpose, the bam file resulting from the back-mapping analysis was converted to a blobtools readable cov file with *blobtools map2cov*. Taxonomic

assignment for BlobTools was conducted with blastn v.2.10.0+ (Camacho et al. 2009) using -task megablast and -e-value 1e-25. The blobDB was created and plotted with the cov file and blast hits. The nuclear draft genome assembly was deposited in GenBank under accession JAPMAE000000000. The DNA barcode region was extracted from the genome assembly and aligned to the traditionally sequenced mtCOI sequences and showed 100% identity to the larvae included by Engelhardt et al. (2011).

Genomic methods were identical for *R. tsurakiana* (see Suppl. material 3).

Morphological examination

The holotype specimen was prepared and examined following standard methods for ethanol-preserved material (Blahnik and Holzenthal 2004; Blahnik et al. 2007). Forewing length was measured from base to apex with a microscale (BioQuip Products, Rancho Dominguez, California, USA).

The abdomen was removed from the specimen, soaked in 85% lactic acid, and heated to 99 °C for 60 min to dissolve internal soft tissues. The macerated tissues were then flushed out of the abdomen with a syringe. The holotype was examined on an Olympus SZX10 stereomicroscope, and pencil sketches of the genitalia were prepared using a Leitz Dialux 20 compound microscope outfitted with a drawing tube. Pencil sketches were scanned and placed in an Adobe Illustrator CS6 document as a template for vector illustrations. Morphological terminology follows Schmid (1970) for the male genitalia, Holzenthal et al. (2007) for wing venation, and Ivanov (1990) for setal warts.

Results

Phylogenetic analysis

After collapsing clades with less than 70% bootstrap support, the species in the *Rhyacophila tristis* group were placed in a polytomy, and sister to *R. italica* (Fig. 2). The first clade of the polytomy includes *R. pubescens*, *R. tsurakiana*, and the new species. The holotype specimen (marked with an asterisk in Fig. 2) was included in a highly supported clade (100% bootstrap support) with three *Rhyacophila* larvae from Corsica. Based solely on larval identification, these were originally considered to be *R. pubescens* (Engelhardt 2009; Engelhardt et al. 2011). It now seems clear that these are larvae of *R. lignumvallis* sp. nov.

The second clade in the polytomy includes several species from the *tristis* subgroup (*R. orghidani* Botosaneanu, 1952, *R. cibinensis* Botosaneanu & Marinkovic-Gospodnetic, 1967, *R. margaritae* Kumanski, 1998, *R. bosnica*, and *R. obtusa* Klapalek, 1894), and two species from the *pubescens* subgroup (*R. pirinica* Kumanski, 1980 and *R. producta* McLachlan, 1879). These specimens were collected in Albania, Austria, Bulgaria, and Romania. The third clade in the polytomy exclusively includes species from the *tristis* subgroup (*R. aquitanica* McLachlan, 1879, *R. carpathica* Botosaneanu, 1995, *R. trescavicensis* Botosaneanu, 1960, *R. tristis* Pictet, 1834, *R. vranitzensis* Botosaneanu & Marinkovic-Gospodnetic, 1967, and *Rhyacophila* sp.). These specimens were collected in Austria, Bosnia and Herzegovina, Bulgaria, France, Italy, Montenegro, Romania, and Spain.



Figure 1. Type locality of *Rhyacophila lignumvallis* sp. nov. on the island of Corsica (France).

Whole genome sequencing and genome characterization of *R. lignumvallis* sp. nov.

Illumina sequencing resulted in 212,866,450 raw reads with a data amount of 31.9 Gbp for *R. lignumvallis* sp. nov. After trimming and contamination filtering, 173,132,236 reads (22.2 Gbp) were kept. The Genomescope2 analysis revealed a genome size of 699,853,381 bp and heterozygosity of 20% (see Suppl. material 2).

The NOVOplasty mitogenome assembly resulted in three contigs (18,087 bp, 1,404 bp, 238 bp) that could not be circularized. Therefore, the 15,623 bp long contig obtained by MitoZ was chosen for annotation. The annotation of the mitogenome revealed all expected 13 protein-coding genes and both rRNAs and 23 tRNAs.

The nuclear genome assembly of *R. lignumvallis* sp. nov. contains 206,802 scaffolds with a total length of 644 Mb, an N50 of 5.6 kb, and a GC of 30%. The BUSCO search with 2,124 Endopterygota orthologs resulted in 82.5% BUSCOs; of these, 47.9% were complete (47.4% single, 0.5% duplicated), and 34.6% were fragmented. 96.3% of the reads were mapped back to the original assembly. Blobtools detected no contamination in the assembly for *R. lignumvallis* sp. nov. (see Suppl. material 2). However, some contamination was detected by NCBI using the improved FCS-GX screen according to <https://github.com/ncbi/fcs> (see Suppl. material 2).

The genomic characterization of *R. tsurakiana* is included in Suppl. material 3.



Figure 2. Phylogenetic relationships in the *Rhyacophila tristis* group based on the mtCOI barcode region and estimated by maximum likelihood, as implemented in IQ-TREE v.2.1.3 (Minh et al. 2020). Nodal support was calculated using the ultrafast bootstrap (UFboot) approximation (Hoang et al. 2017); nodes with UFboot values greater than 95% are considered well supported. Branches with support values of less than 70% were collapsed. Branch lengths are measured in nucleotide substitutions per site (see scale bar for reference). The branch leading to *Rhyacophila* was shortened (total branch length was 0.12 substitutions per site). Numbers in parentheses correspond to GenBank accession numbers. The holotype and the Corsican larvae are shaded.

Species description

Rhyacophila lignumvallis Graf & Rázuri-Gonzales, sp. nov.

<https://zoobank.org/5A58EBFA-E945-4032-917A-877444F5DA2B>

Holotype. FRANCE • ♂; Corsica, bridge over the river Vecchio near the confluence with the river Tavignano; 42.2275°N, 9.24306°E; 195 m a.s.l.; 25 Jul. 2019; col. W. Graf leg.; in ethanol; SMF (SMFTRI00018634).

Diagnosis. *Rhyacophila lignumvallis* sp. nov. (Figs 4, 5A, B) belongs to the *Rhyacophila tristis* species group (Schmid 1970). It is most similar to *R. pubescens* (Fig. 5C, D), *R. tsurakiana* (Fig. 5E, F), *R. ligurica* Oláh & Vinçon, 2021 (in Oláh et al. 2021, figs 55–57 therein), *R. harmasa* Oláh & Vinçon, 2021 (in Oláh et al. 2021, figs 52–54 therein), and *R. abruzzica* Oláh & Vinçon, 2021 (in Oláh et al. 2021, figs 49–51 therein), but *R. lignumvallis* sp. nov. is distinguishable from these species by the shape of tergum X, the dorsal arm of the phallic apparatus in lateral and ventral views, the shape of the aedeagus and parameres, and the second segment of the inferior appendages.

The dorsal surface of segment X is convex in all these species but narrower and higher in the new species, *R. tsurakiana*, and *R. harmasa*. In dorsal view, however, the new species has a slightly membranous, mesally notched, and inflated segment X, while segment X in *R. tsurakiana* appears flatter. Additionally, the dorsal branch of tergum X is rounded and broader in the new species, while it is narrower in *R. tsurakiana* and *R. harmasa*.

The dorsal appendix of the phallic apparatus in the new species is longer than the aedeagus and the parameres (Fig. 4D). This also occurs in *R. harmasa*, *R. ligurica*, *R. pubescens*, and *R. tsurakiana* but not in *R. abruzzica*. However, the shape of the dorsal appendix in lateral view in the new species is digitate and slightly curved dorsad, whereas *R. harmasa* has a slightly wider apical half, *R. ligurica* has a low bump mesally on its dorsal surface, and *R. tsurakiana* has a straight and flat dorsal appendix. In *R. abruzzica*, the dorsal appendage is broad and medially widened in lateral view. In comparison to *R. lignumvallis* sp. nov. (Figs 4D, 5A, B), *R. tsurakiana*, and *R. abruzzica*, the dorsal appendix of the phallic apparatus is much longer and clearly exceeds segment X in dorsal view in *R. harmasa*, *R. pubescens*, and *R. ligurica*. In ventral view, the dorsal appendix is straight and rounded apically in the new species (Fig. 4E), slightly inflated on the apical half and rounded apically in *R. harmasa*, almost straight and truncate apically in *R. ligurica*, constricted basally and truncate apically in *R. tsurakiana*, and rectangular in *R. abruzzica*.

The aedeagus and parameres in *R. lignumvallis* sp. nov. are most similar to *R. pubescens*. However, in lateral view, the tip of the aedeagus in the new species is slenderer and slightly more curved apically than in *R. pubescens*. In lateral view, the parameres in the new species are broader than in *R. pubescens*. In ventral view, the parameres in the new species are club-shaped and curved mesad, while in *R. pubescens*, the parameres are digitate and directed posterad.

The second segment of the inferior appendages in the new species is triangular, with a straight dorsal margin, while all the other species have a concave dorsal margin (Fig. 4A, 5B).

Description. Adult male. Specimen in ethanol, mostly denuded; dorsally brown, ventrally light brown. Legs light brown with slightly darker tibial spurs. Head with frontal setal wart triangular; antennal setal wart subtriangular and smaller than frontal setal wart; posterior setal warts oval and connected to ocellar setal warts via a raised cuticular “bridge” (see Schmid, 1970; pl. I, fig. 1). Forewing length (8.8 mm, $N = 1$) mostly denuded, with sparse, very short light brown setae and golden brown microtrichia. Hind wings also mostly denuded, with slightly longer light brown setae. Forewing (Fig. 3A) with crossveins connecting costal (C) and subcostal (Sc) veins; subcostal (Sc) and first radial (R_1) veins, first (R_1) and second radial (R_2) veins (r), fifth radial (R_5) and first medial (M) veins ($r-m$), first medial (M) and first cubital (Cu1) veins ($m-cu$), and first cubital (Cu1) and second cubital (Cu2) veins. Hind wing (Fig. 3B) with crossveins connecting subcostal (Sc) and first radial (R_1) veins, fifth radial (R_5) and first medial (M) veins ($r-m$), and M_{3+4} and first cubital (Cu1) present.

Male genitalia. Segment IX longitudinally short in lateral view (Fig. 4A), anterior and posterior margins slightly concave, dorsal half slightly longer than ventral. Dorsal surface of segment X membranous, slightly inflated, shallowly notched mesally in dorsal view (Fig. 4C). Dorsal branch of segment X short and rounded in lateral view. Anal sclerites partially fused to each other basally and to segment X, in lateral view, directed ventrad. First article of inferior appendages (Fig. 4A) rect-

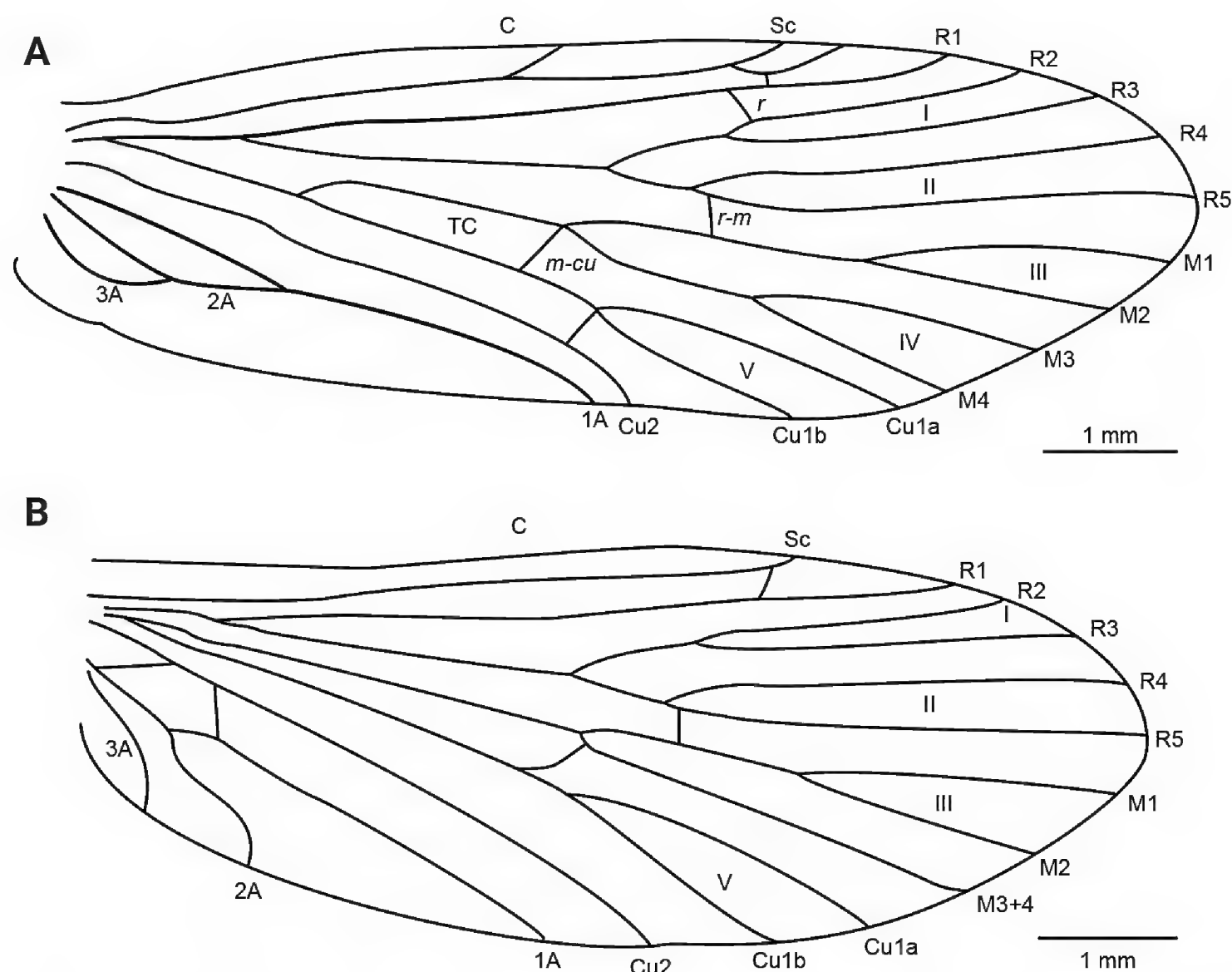


Figure 3. *Rhyacophila lignumvallis* sp. nov., wing venation. **C**, costal vein; **Sc**, subcostal vein; **R1–R5**, first to fifth branches of the radial vein; **M1–M4**, first to fourth branches of the medial vein; **M3+4**, medial vein 3+4 (hindwing); **Cu1a**, anterior branch of first cubital vein; **Cu1b**, posterior branch of first cubital vein; **Cu2**, second cubital vein; **1A–3A**, first to third anal veins; **r**, radial crossvein; **r-m**, radiomedial crossvein; **m-cu**, mediocubital crossvein; **I–V**, first to fifth wing forks; **TC**, thyridial cell. Scale bar: 1 mm. Illustrations were produced by Ernesto Rázuri-Gonzales.

angular in lateral view, slightly broader basally than apically; in ventral view (Fig. 4B), slightly broader apically than basally, with a small setose bump basally on mesal surface. Second article of inferior appendages (Fig. 4A) quadrangular in lateral view, dorsal and ventral margins slightly diverging, posterodorsal margin straight, at a 130° angle to dorsal margin; in ventral view (Fig. 4B), mitten-shaped, mesal margin with very short, peg-like setae basally and longer setae apically. Phallic apparatus (Fig. 4D) with dorsal appendix straight in lateral view, slightly curved dorsad, rounded apically, longer than parameres and aedeagus; in ventral view, straight, lateral margins slightly sinuous, rounded apically. Parameres in lateral view (Fig. 4D) broader than aedeagus, slightly curved posterodorsad, ventral margin straight, dorsal margin slightly sinuous, rounded apically; in ventral view (Fig. 4E), club-shaped, directed mesad. Aedeagus in lateral view (Fig. 4D) slender, slightly sinuous, tapering towards its apex; in ventral view, slender and straight.

Etymology. We dedicate this species to Dr Ralph W. Holzenthal to honor his contributions to caddisfly taxonomy and systematics. *Lignumvallis*, wood valley, is derived from the Latin translation of Ralph's last name.

Habitat. The river Vecchio is a crystal-clear, slow-flowing stream with a heterogeneous bottom substrate that varies from sandy patches to gravel to boulders. Stable substrates were densely covered by *Agapetus cyrnensis* pupae. As many spring trickles enter the river on its left margin and the specimen was collected by sweeping the vegetation, the habitat of *R. lignumvallis* sp. nov. remains unknown.

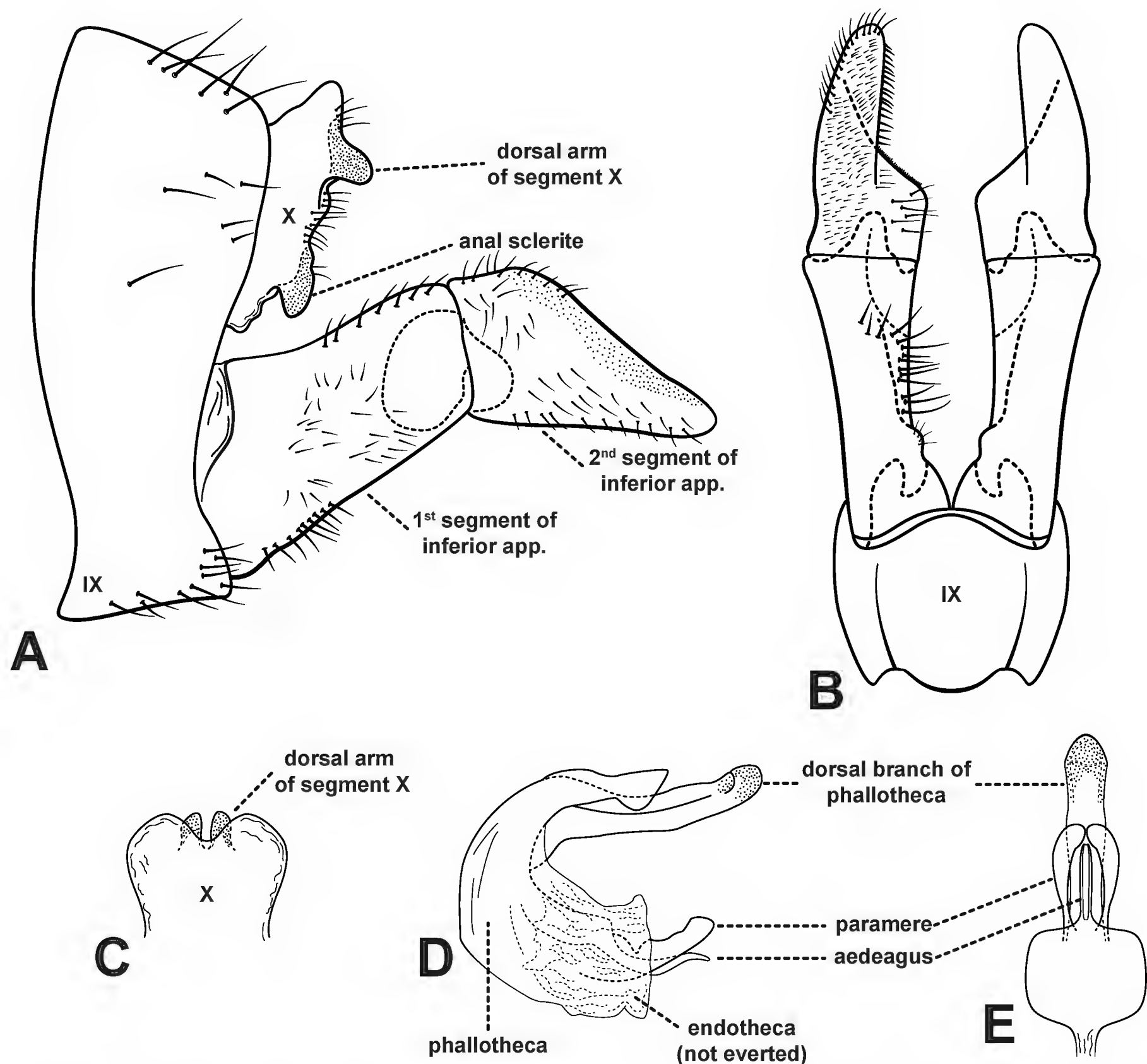


Figure 4. *Rhyacophila lignumvallis* sp. nov., male genitalia, lateral view (A), ventral view (B), segment X in dorsal view (C), phallic apparatus in lateral view (D), and phallic apparatus in ventral view (E). Illustrations were produced by Ernesto Rázuri-Gonzales.

Discussion

Despite being one of the most well-known faunas in the world, on average more than 770 new animal species are described from Europe each year (Fontaine et al. 2012), with many taxonomic groups not yet reaching a plateau (e.g., Mateos et al. 2017). This trend is particularly pronounced among endemic species, with conservative estimates suggesting that up to one-fifth of endemic taxa from Europe have not yet been described (Essl et al. 2013). Many of these narrowly distributed species are characterized by inhabiting very small, isolated habitats and generally having small populations, making them especially vulnerable to environmental changes (Hering et al. 2009; Essl et al. 2013). This vulnerability is augmented in species occurring at higher elevations that may be subject to “summit trap effects” (Bálint et al. 2011; Domisch et al. 2011; Sauer et al. 2011; Taubmann et al. 2011). Discovering and potentially safeguarding these species prior to their extirpation should be a priority in conservation efforts.

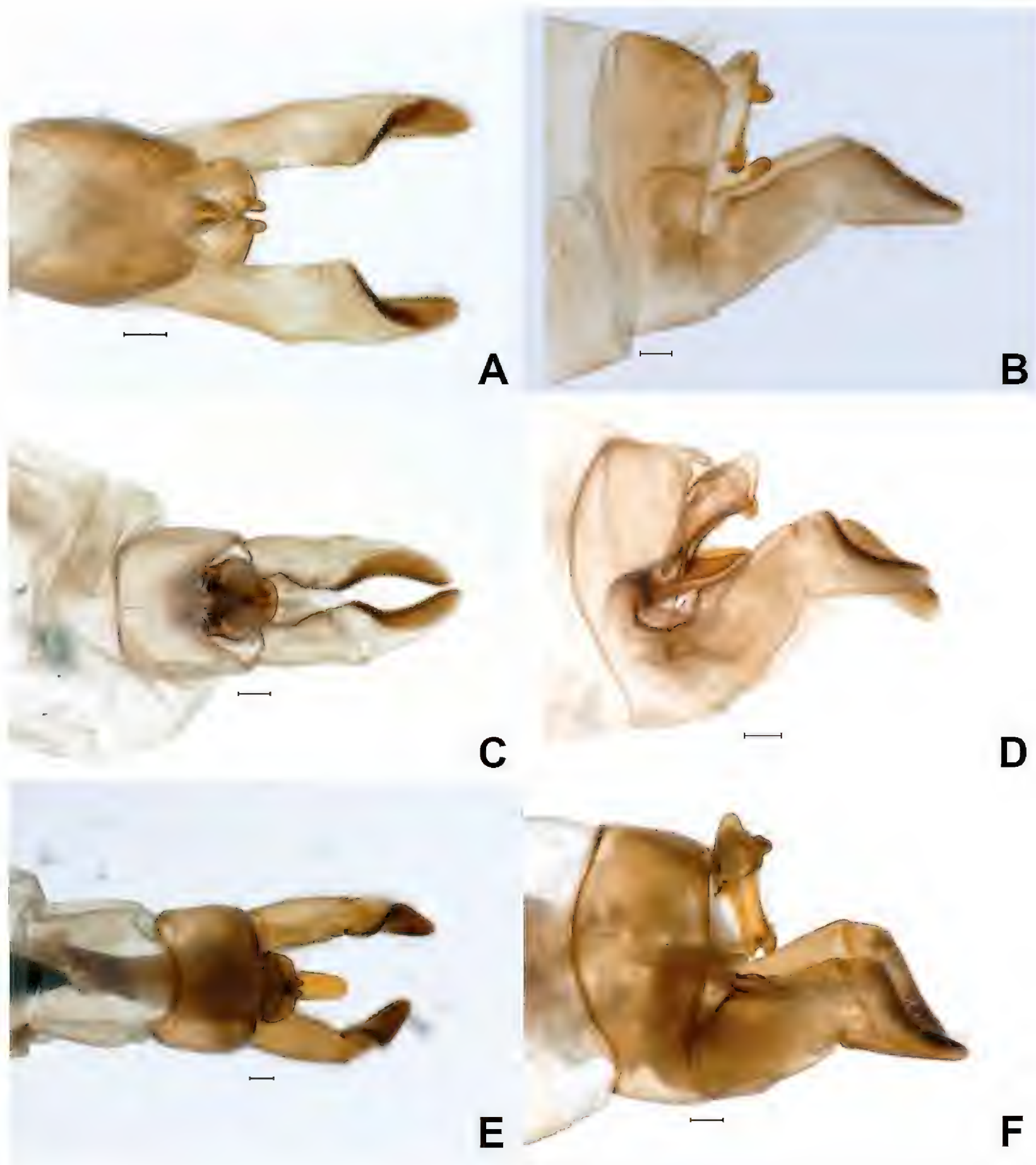


Figure 5. *Rhyacophila lignumvallis* sp. nov., male genitalia, dorsal view (A), lateral view (B). *R. pubescens*, male genitalia, dorsal view (C), lateral view (D). *R. tsurakiana*, male genitalia, dorsal view (E), lateral view (F). Scale bars: 100 µm. Specimens were photographed by W. Graf.

In the case of caddisflies, southern Europe (e.g., Spain, Italy, and the Balkan Peninsula), mountainous regions (e.g., the Alps and the Pyrenees), and the Caucasus have been shown to be particularly species-rich and centers of endemism (Pauls et al. 2006; Previšić et al. 2014; Graf et al. 2015; Schmidt-Kloiber et al. 2017). Although caddisflies are well-studied in most of Europe, further studies in these highly diverse areas are necessary to better understand their richness in this continent and will likely yield many more new species.

The *Rhyacophila tristis* group now includes 29 species distributed throughout central-southern Europe and Western Asia, with many of them occurring in biodiversity centers in these regions (e.g., southern Europe and the Balkans, Suppl. material 1). Additionally, many species in this group are only known from

a single locality, a single or very few specimens, or with unknown females/immature stages. For example, the single adult specimen of *R. lignumvallis* sp. nov. was associated with larval specimens from Corsica, previously identified as *R. pubescens* (Engelhardt 2009). Further sampling will clarify the potential presence of *R. pubescens* on the island. This suggests that the taxonomy of this group is far from complete, particularly for juvenile stages.

Aquatic insects have traditionally been neglected in genomic research (Hotaling et al. 2020). Using 40× coverage of short-read sequencing, we were able to obtain a draft nuclear and complete mitogenome assembly for the holotype of *R. lignumvallis* sp. nov. The assembly of the newly described species was 644,010,216 bp in length, which is close to the estimate obtained by Genomescope2. With an N50 of 5.6 Kbp, the genome assembly is less contiguous than previously published *Rhyacophila* genomes (*R. brunnea* Banks, 1911 and *R. evoluta* McLachlan, 1879 in Heckenhauer et al. (2023)). This lower contiguity is probably due to the sequencing technologies used (Oxford Nanopore long-reads followed by polishing with Illumina short-reads for the *R. brunnea* genome assembly vs. Illumina short-reads only for the *R. lignumvallis* sp. nov. genome) and/or sequencing coverage (97× and 116× Illumina filtered reads for the two *R. evoluta* genome assemblies vs. 40× in the new species) (Heckenhauer et al. 2022).

The percentage of BUSCOs recovered in the draft genome assembly was 82.5%. Of these, 47.9% were complete and 34.6% were fragmented. Meanwhile, the previously generated *Rhyacophila* genomes had a complete BUSCO score of 95.4% for *R. brunnea* (only 2.5% fragmented) and 74.1/75.1% for two *R. evoluta* specimens (17.9/18.7% fragmented) (Heckenhauer et al. 2022). The discrepancy in the amount of complete and fragmented BUSCOs among these congeners is probably due to the reasons outlined for the contiguity.

Genome assembly quality can be assessed using various metrics, such as contiguity and BUSCO completeness (Gurevich et al. 2013; Heckenhauer et al. 2022). Clearly, this genomic characterization is far from a reference genome quality, but nevertheless permanently connects the species name, the underlying morphology as preserved in the type specimen with the genetic make-up of the most representative specimen of the species (e.g., Hebert and Gregory 2005; Padial and de la Riva 2007; Pohl et al. 2012; Egan et al. 2017; Heckenhauer et al. 2023). This information is valuable for studying the systematics and evolution of the species in question as described in Heckenhauer et al. (2023). Additionally, the complete mitogenome includes the DNA barcode, which has already become important to species descriptions and can be used to monitor the occurrence of this species in freshwater bodies.

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Additional information

Conflict of interest

The authors have declared that no competing interests exist.

Ethical statement

No ethical statement was reported.

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Author contributions

ER: investigation, data curation, formal analysis, writing – original draft, writing – review and editing, visualization. WG: investigation, writing – original draft, writing – review and editing, visualization. JH: data curation, formal analysis, writing – original draft, writing – review and editing. JS: investigation, writing – original draft, writing – review and editing. SUP: study design, data curation, investigation, writing – original draft, writing – review and editing.

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Data availability

All of the data that support the findings of this study are available in the main text or Supplementary Information.

References

- Andrews S (2019) FastQC: a quality control tool for high throughput sequence data. v. 0.11.9, <http://www.bioinformatics.babraham.ac.uk/projects/fastqc/>
- Bálint M, Domisch S, Engelhardt CHM, Haase P, Lehrian S, Sauer J, Theissinger K, Pauls SU, Nowak C (2011) Cryptic biodiversity loss linked to global climate change. *Nature Climate Change* 1: 313–318. <https://doi.org/10.1038/nclimate1191>
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA (2012) SPAdes: A New Genome Assembly Algorithm and Its Applications to Single-Cell Sequencing. *Journal of Computational Biology* 19: 455–477. <https://doi.org/10.1089/cmb.2012.0021>
- Banks N (1911) Description of new species of North American neuropteroid insects. *Transactions of the American Entomological Society* 37: 335–360, plates 311–313.
- Bianchini G, Sánchez-Baracaldo P (2024) TreeViewer: Flexible, modular software to visualise and manipulate phylogenetic trees. *Ecology and Evolution* 14: e10873. <https://doi.org/10.1002/ece3.10873>

- Blahnik RJ, Holzenthal RW (2004) Collection and curation of Trichoptera, with an emphasis on pinned material. Nectopsyche, Neotropical Trichoptera Newsletter 1: 8–20. <http://hdl.handle.net/11299/190744>
- Blahnik RJ, Holzenthal RW, Prather AL (2007) The lactic acid method for clearing Trichoptera genitalia. In: Bueno-Soria J, Barba-Álvarez R, Armitage BJ (Eds) Proceedings of the 12th International Symposium on Trichoptera. The Caddis Press, Columbus, Ohio, 9–14.
- Bolger AM, Lohse M, Usadel B (2014) Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30: 2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>
- Botosaneanu L (1952) *Rhyacophila orghidani* n. sp. (Trichoptera Rhyacophilinae) din Muntii Apuseni ai Republicii Populare Române. Comunicările Academiei Republicii Populare Române 2: 721–724.
- Botosaneanu L (1960) Trichoptères de Yougoslavie recueillis en 1955 par le Dr. F. Schmid. Deutsche Entomologische Zeitschrift 7: 261–293 <https://doi.org/10.1002/mmnd.19600070304>
- Botosaneanu L (1995) Additional documents to the knowledge of the Trichoptera of Romania, with data on European taxa from outside this country (Insecta: Trichoptera). Faunistische Abhandlungen Staatliches Museum für Tierkunde Dresden 20: 57–88.
- Botosaneanu L, Marinkovic-Gospodnetic M (1967) Sur quelques *Rhyacophila* du groupe de *tristis* (Trichoptera). Annales de la Société Entomologique de France (NS) 3: 1145–1151.
- Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL (2009) BLAST+: architecture and applications. BMC Bioinformatics 10: 421. <https://doi.org/10.1186/1471-2105-10-421>
- Coppa G, Graf W, Tachet H (2012) A revised description of the larvae of three species of the *Rhyacophila tristis* group: *Rhyacophila aquitanica*, *Rhyacophila pubescens* and *Rhyacophila tristis* (Trichoptera: Rhyacophilidae). Annales de Limnologie 48: 215–223. <https://doi.org/10.1051/limn/2012014>
- Dierckxsens N, Mardulyn P, Smits G (2016) NOVOPlasty: de novo assembly of organelle genomes from whole genome data. Nucleic Acids Research 45: e18. <https://doi.org/10.1093/nar/gkw955>
- Döhler W (1950) Zur Kenntnis der Gattung *Rhyacophila* im mitteleuropäischen Raum (Trichoptera). Archiv für Hydrobiologie 44: 271–293.
- Domisch S, Jähnig SC, Haase P (2011) Climate-change winners and losers: stream macroinvertebrates of a submontane region in Central Europe. Freshwater Biology 56: 2009–2020. <https://doi.org/10.1111/j.1365-2427.2011.02631.x>
- Egan SP, Weinersmith KL, Liu S, Ridenbaugh RD, Zhang YM, Forbes AA (2017) Description of a new species of *Euderus* Haliday from the southeastern United States (Hymenoptera, Chalcidoidea, Eulophidae): the crypt-keeper wasp. ZooKeys 645: 37–49. <https://doi.org/10.3897/zookeys.645.11117>
- Engelhardt C (2009) Phylogeny and phylogeography of the caddisfly *Rhyacophila pubescens*, PICTET 1834, (Trichoptera), with special consideration of its habitat specificity. PhD Thesis, Universität Duisburg-Essen, Essen, 120 pp.
- Engelhardt CHM, Haase P, Pauls SU (2011) From the Western Alps across Central Europe: postglacial recolonisation of the tufa stream specialist *Rhyacophila pubescens* (Insecta, Trichoptera). Frontiers in Zoology 8: 10 [14 pp]. <https://doi.org/10.1186/1742-9994-8-10>
- Essl F, Rabitsch W, Dullinger S, Moser D, Milasowszky N (2013) How well do we know species richness in a well-known continent? Temporal patterns of endemic and widespread species descriptions in the European fauna. Global Ecology and Biogeography 22: 29–39. <https://doi.org/10.1111/j.1466-8238.2012.00787.x>

- Fontaine B, van Achterberg K, Alonso-Zarazaga MA, Araujo R, Asche M, Aspöck H, Aspöck U, Audisio P, Aukema B, Bailly N, Balsamo M, Bank RA, Belfiore C, Bogdanowicz W, Boxshall G, Burckhardt D, Chylarecki P, Deharveng L, Dubois A, Enghoff H, Fochetti R, Fontaine C, Gargominy O, Lopez MSG, Goujet D, Harvey MS, Heller K-G, van Helsdingen P, Hoch H, De Jong Y, Karsholt O, Los W, Magowski W, Massard JA, McInnes SJ, Mendes LF, Mey E, Michelsen V, Minelli A, Nafria JMN, van Nieukerken EJ, Pape T, De Prins W, Ramos M, Ricci C, Roselaar C, Rota E, Segers H, Timm T, van Tol J, Bouchet P (2012) New species in the old world: Europe as a frontier in biodiversity exploration, a test bed for 21st century taxonomy. *PLOS ONE* 7: e36881. <https://doi.org/10.1371/journal.pone.0036881>
- Graf W, Vitecek S, Previsic A, Malicky H (2015) New species of Limnephilidae (Insecta: Trichoptera) from Europe: Alps and Pyrenees as harbours of unknown biodiversity. *Zootaxa* 3911: 381–395. <https://doi.org/10.11646/zootaxa.3911.3.5>
- Gurevich A, Saveliev V, Vyahhi N, Tesler G (2013) QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 29: 1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>
- Hagen HA (1859) Die Phryganiden Pictet's nach typen bearbeitet (continued in 1860 and 1861). *Stettiner Entomologische Zeitung* 20: 131–170.
- Hebert PDN, Gregory TR (2005) The Promise of DNA Barcoding for Taxonomy. *Systematic Biology* 54: 852–859. <https://doi.org/10.1080/10635150500354886>
- Heckenhauer J, Frandsen PB, Sproul JS, Li Z, Paule J, Larracuente AM, Maughan PJ, Barker MS, Schneider JV, Stewart RJ, Pauls SU (2022) Genome size evolution in the diverse insect order Trichoptera. *GigaScience* 11. <https://doi.org/10.1093/gigascience/giac011>
- Heckenhauer J, Rázuri-Gonzales E, Mwangi FN, Schneider J, Pauls SU (2023) Holotype sequencing of *Silvatares holzenthali* Rázuri-Gonzales, Ngera & Pauls, 2022 (Trichoptera, Pisuliidae). *ZooKeys* 1159. <https://doi.org/10.3897/zookeys.1159.98439>
- Hering D, Schmidt-Kloiber A, Murphy J, Lücke S, Zamora-Muñoz C, López-Rodríguez MJ, Huber T, Graf W (2009) Potential impact of climate change on aquatic insects: a sensitivity analysis for European caddisflies (Trichoptera) based on distribution patterns and ecological preferences. *Aquatic Sciences* 71: 3–14. <https://doi.org/10.1007/s00027-009-9159-5>
- Hoang DT, Chernomor O, von Haeseler A, Minh BQ, Vinh LS (2017) UFBoot2: Improving the Ultrafast Bootstrap Approximation. *Molecular Biology and Evolution* 35: 518–522. <https://doi.org/10.1093/molbev/msx281>
- Holzenthal RW, Blahnik RJ, Prather AL, Kjer KM (2007) Order Trichoptera Kirby, 1813 (Insecta), caddisflies. *Zootaxa* 1668: 639–698. <https://doi.org/10.11646/zootaxa.1668.1.29>
- Hotaling S, Kelley JL, Frandsen PB (2020) Aquatic Insects Are Dramatically Underrepresented in Genomic Research. *Insects* 11: 601. <https://doi.org/10.3390/insects11090601>
- Hwang C-L (1958) Descriptions of Chinese caddis flies (Trichoptera). *Acta Entomologica Sinica* 10: 279–285.
- Ivanov VD (1990) Structure and function of setose warts of caddisflies [in Russian]. *Latvijas Entomologs* 33: 96–110.
- Kalyaanamoorthy S, Minh BQ, Wong TKF, von Haeseler A, Jermiin LS (2017) ModelFinder: fast model selection for accurate phylogenetic estimates. *Nature Methods* 14: 587–589. <https://doi.org/10.1038/nmeth.4285>
- Klapálek F (1894) Descriptions of new species of *Raphidia*, L., and of three new species of Trichoptera from the Balkan Peninsula, with critical remarks on *Panorpa gibberosa*, McLachlan. *Transactions of the Entomological Society of London* 1894: 489, 495. <https://doi.org/10.1111/j.1365-2311.1894.tb02097.x>

- Kumanski KP (1980) Description of three new caddis-flies (Trichoptera) from Bulgaria. *Rivista di Idrobiologia* 19: 197–206.
- Kumanski KP (1998) *Rhyacophila margaritae* – a new insect species (Trichoptera: Rhyacophilidae) from Bulgaria. *Dokladi na Bulgarskata Akademiya na Naukite* 51: 59–62.
- Laetsch DR, Blaxter ML (2017) BlobTools: Interrogation of genome assemblies. *F1000Research* 6. <https://doi.org/10.12688/f1000research.12232.1>
- Li H (2013) Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. *arXiv [q-Bio.GN]*. <https://doi.org/10.48550/arXiv.1303.3997>
- Malicky H (1984) Fünf neue griechische Köcherfliegen (Trichoptera). *Mitteilungen der Entomologischen Gesellschaft Basel* 34: 96–102.
- Manni M, Berkeley MR, Seppey M, Zdobnov EM (2021) BUSCO: Assessing Genomic Data Quality and Beyond. *Current Protocols* 1: e323. <https://doi.org/10.1002/cpz1.323>
- Marçais G, Kingsford C (2011) A fast, lock-free approach for efficient parallel counting of occurrences of k-mers. *Bioinformatics* 27: 764–770. <https://doi.org/10.1093/bioinformatics/btr011>
- Mateos E, Sluys R, Riutort M, Álvarez-Presas M (2017) Species richness in the genus *Microplana* (Platyhelminthes, Tricladida, Microplaninae) in Europe: as yet no asymptote in sight. *Invertebrate Systematics* 31: 269–301. <https://doi.org/10.1071/IS16038>
- McLachlan R (1879) A monographic revision and synopsis of the Trichoptera of the European fauna. Part 8. John van Voorst, London, 429–500, plates 445–451 pp.
- Meng G, Li Y, Yang C, Liu S (2019) MitoZ: a toolkit for animal mitochondrial genome assembly, annotation and visualization. *Nucleic Acids Research* 47: e63. <https://doi.org/10.1093/nar/gkz173>
- Minh BQ, Schmidt HA, Chernomor O, Schrempf D, Woodhams MD, von Haeseler A, Lanfear R (2020) IQ-TREE 2: New Models and Efficient Methods for Phylogenetic Inference in the Genomic Era. *Molecular Biology and Evolution* 37: 1530–1534. <https://doi.org/10.1093/molbev/msaa015>
- Moretti GP (1981) New Trichoptera species and subspecies found in Italy. In: Moretti GP (Ed.) *Proceedings of the 3rd International Symposium on Trichoptera*. Dr. W. Junk, The Hague, 165–192. https://doi.org/10.1007/978-94-009-8641-1_22
- Mosely ME (1930) Corsican Trichoptera. *Eos – Revista Española Entomología* 6: 147–184.
- Mosely ME (1932) Corsican Trichoptera and Neuroptera (s. l.). *Eos – Revista Española de Entomología* 8: 165–184.
- Nelson LA, Wallman JF, Downton M (2007) Using COI barcodes to identify forensically and medically important blowflies. *Medical and Veterinary Entomology* 21: 44–52. <https://doi.org/10.1111/j.1365-2915.2007.00664.x>
- Okonechnikov K, Conesa A, García-Alcalde F (2016) Qualimap 2: advanced multi-sample quality control for high-throughput sequencing data. *Bioinformatics* 32: 292–294. <https://doi.org/10.1093/bioinformatics/btv566>
- Oláh J, Vinçon G, Coppa G (2021) On the Trichoptera of Italy with delineation of incipient sibling species. *Opuscula Zoologica (Budapest)* 52: 03–67. <https://doi.org/10.18348/opzool.2021.1.3>
- Oláh J, Beshkov S, Ibrahim H, Kovács T, Oláh Jr J, Vinçon G (2022) On the Trichoptera of the Balkan: survey of species complexes of *Polycentropus ierapetra*, *Rhyacophila balcanica*, *R. bosnica* and *Notidobia nekibe*. *Opuscula Zoologica (Budapest)* 53: 67–111. <https://doi.org/10.18348/opzool.2022.1.67>
- Padial JM, de la Riva I (2007) Integrative taxonomists should use and produce DNA barcodes. *Zootaxa* 1586: 67–68. <https://doi.org/10.11646/zootaxa.1586.1.7>

- Pauls SU, Lumbsch HT, Haase P (2006) Phylogeography of the montane caddisfly *Drusus discolor*: evidence for multiple refugia and periglacial survival. *Molecular Ecology* 15: 2153–2169. <https://doi.org/10.1111/j.1365-294X.2006.02916.x>
- Pictet FJ (1834) Recherches pour servir à l'histoire et l'anatomie des Phryganides. A. Cherbuliez, Geneva, plates 1–20 + 235 [220 plates]. <https://doi.org/10.5962/bhl.title.8547>
- Pohl H, Niehuis O, Gloyna K, Misof B, Beutel R (2012) A new species of *Mengenilla* (Insecta, Strepsiptera) from Tunisia. *ZooKeys* 198: 79–102. <https://doi.org/10.3897/zookeys.198.2334>
- Previšić A, Graf W, Vitecek S, Kučinić M, Bálint M, Keresztes L, Pauls SU, Waringer J (2014) Cryptic diversity of caddisflies in the Balkans: the curious case of *Ecclisopteryx* species (Trichoptera: Limnephilidae). *Arthropod Systematics and Phylogeny* 72: 309–329. <https://doi.org/10.3897/asp.72.e31792>
- Ranallo-Benavidez TR, Jaron KS, Schatz MC (2020) GenomeScope 2.0 and Smudgeplot for reference-free profiling of polyploid genomes. *Nature Communications* 11: 1432. <https://doi.org/10.1038/s41467-020-14998-3>
- Rohland N, Reich D (2012) Cost-effective, high-throughput DNA sequencing libraries for multiplexed target capture. *Genome Research* 22: 939–946. <https://doi.org/10.1101/gr.128124.111>
- Ross HH (1956) Evolution and Classification of the Mountain Caddisflies. University of Illinois Press, Urbana, 213 pp.
- Sauer J, Domisch S, Nowak C, Haase P (2011) Low mountain ranges: summit traps for montane freshwater species under climate change. *Biodiversity and Conservation* 20: 3133–3146. <https://doi.org/10.1007/s10531-011-0140-y>
- Schmid F (1970) Le genre *Rhyacophila* et la famille des Rhyacophilidae (Trichoptera). *Memoires de la Société Entomologique du Canada* 66: 1–230. <https://doi.org/10.4039/entm10266fv>
- Schmidt-Kloiber A, Neu PJ, Malicky M, Pletterbauer F, Malicky H, Graf W (2017) Aquatic biodiversity in Europe: a unique dataset on the distribution of Trichoptera species with important implications for conservation. *Hydrobiologia* 797: 11–27. <https://doi.org/10.1007/s10750-017-3116-4>
- Sun C-H, Yang L-F (1995) Studies on the genus *Rhyacophila* (Trichoptera) in China (1). *Braueria* 22: 27–32.
- Taubmann J, Theissinger K, Feldheim KA, Laube I, Graf W, Haase P, Johannesen J, Pauls SU (2011) Modelling range shifts and assessing genetic diversity distribution of the montane aquatic mayfly *Ameletus inopinatus* in Europe under climate change scenarios. *Conservation Genetics* 12: 503–515. <https://doi.org/10.1007/s10592-010-0157-x>
- Ulmer G (1927) Einige neue Trichopteren aus Asien. *Entomologische Mitteilungen, Deutsche Entomologische Museum* 16: 172–182.
- Valladolid M, Waringer J, Arauzo M, Chvojka P, Dorda BA, Komzák P, Lodovici O, Rey I (2023) The *Rhyacophila fasciata* Species Complex (Trichoptera: Rhyacophilidae) in Central Europe with description of a new species, *Rhyacophila loeffleri* Valladolid & Waringer, n. sp., based on morphological, genetic and ecological evidence. *Zootaxa* 5325: 451–482. <https://doi.org/10.11646/zootaxa.5325.4.1>
- Waldvogel A-M, Wieser A, Schell T, Patel S, Schmidt H, Hankeln T, Feldmeyer B, Pfenniger M (2018) The genomic footprint of climate adaptation in *Chironomus riparius*. *Molecular Ecology* 27: 1439–1456. <https://doi.org/10.1111/mec.14543>
- Wood DE, Salzberg SL (2014) Kraken: ultrafast metagenomic sequence classification using exact alignments. *Genome Biology* 15: R46. <https://doi.org/10.1186/gb-2014-15-3-r46>

Supplementary material 1

Blobtools graphs for the *Rhyacophila lignumvalle* sp. nov. assembly

Authors: Ernesto Rázuri-Gonzales, Wolfram Graf, Jacqueline Heckenhauer, Julio V. Schneider, Steffen U. Pauls

Data type: docx

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Supplementary material 2

Genomic methods and characterization for the *Rhyacophila tsurakiana* genome assembly

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Data type: docx

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Supplementary material 3

Geographic distribution of species in the *Rhyacophila tristis* species group

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Data type: docx

Explanation note: This list includes the geographic distribution of the 25 currently known species in the *Rhyacophila tristis* species group based on literature records. We only include papers with geographic data.

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